Prospects

The rapeutic potential of targeting Wnt/ β -catenin pathway in treatment of colorectal cancer: rational and progress[†]

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Received 19 January 2017; Accepted 19 January 2017 Journal of Cellular Biochemistry This article is protected by copyright. All rights reserved DOI 10.1002/jcb.25903 Wnt/β-catenin pathway is one of the main/frequent dysregulated pathways in several tumor types, including colon cancer. Aberrant activation of this pathway is associated with cell proliferation, invasive behaviors and cell resistance, suggesting its potential value as a therapeutic target in treatment of CRC. Several agents have been developed for targeting of this pathway (e.g, natural agents: curcumin, 3,3-diindolylmethane, phytoestrogen; Synthetic/small Wnt inhibitors: Rofecoxib; PRI-724, CWP232291; and monoclonal antibody against frizzled receptors, Vanituctumab. This review summarizes the current knowledge about the therapeutic potential of targeting Wnt pathway with particular emphasis on preclinical/clinical studies in treatment of colorectal cancer. This article is protected by copyright. All rights reserved

Key word: Colon cancer, Wnt/β -catenin, inhibitor, therapeutic target

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Introduction

Colorectal cancer (CRC) is third leading cause of cancer mortality, which reported over 1.2 million new patients and 600,000 mortalities every year (Hosseini, 2016; Xiao et al., 2016). Progress in combination chemotherapy such as FOLFIRI, XELOX/ CAPOX, FOLFOX, and supportive consumption of therapeutic antibodies against epidermal growth factor receptor (EGFR) and vascular endothelial growth factor (VEGF) have been shown to increase survival time (Watanabe et al., 2015). Although, it is still remained as one of the leading casue of death, supporting the need for detection of novel agents in treatment of CRC.

Recently Wnt/ β -catenin pathway is emerged as promising therapeutic target in CRC, , since it most of the patients with CRC have mutations in at the least one Wnt signaling cascade gene, such as the β -catenin (CTNNB1) and adenomatous polyposis coli (APC) genes. The APC gene was firstly identified as being main causative for the familial adenomatous polyposis (FAP) syndrome (Preisinger et al., 1991) .But later found to be commonly (~80%) mutated in sporadic colorectal cancers and about half of colon cancers with wild-type APC carry mutations in β -catenin (Morin et al., 1997). In this review, we provide an overview on targeting Wnt/ β -catenin signaling pathway as a potential therapeutic approach in treatment of CRC.

Wnt signaling pathway

Wnt pathway is categorized into canonical and non-canonical (β -catenin independent) signaling pathway. In the absence of wnt ligands, β -catenin situates in cell membrane and communicates with E-cadherin, and mediates intercellular adhesion. Usually, free cytoplasm β -catenin is destabilized and damaged by a destruction complex including APC, Axin, casein kinase 1 α (CK1 α) and glycogen synthase kinase-3b (GSK3b). When Wnt cooperates with Frizzled and LRP, Wnt/ β catenin signaling is activated. Both Dishevelled (Dvl) and Axin are undergo to cell membrane recruitment, followed by GSK3b suppression and β -catenin releasing of destruction complex. Wnt permits β -catenin to accumulate and transformed to the nucleus where it binds to various

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transcription factors, such as T-cell factor (TCF) and lymphoid enhancer factor1 (LEF-1). In the involvement of transcriptional co-activators of the β -catenin or TCF4 complex, cAMP response element-binding protein (CREB)-binding protein (CBP). CBP encompass proteins associated with cell differentiation, proliferation, propagation, apoptosis, invasion and angiogenesis(Barker, 2008; Zhan et al., 2016).

Non-canonical Wnt signaling is distinguished by β-catenin-independent mechanisms of signal transduction. Within Wnt/PCP signaling, Wnt ligands bind to ROR-Frizzled receptor and activate Dvl. Then, Dvl binds to the small GTPase Rho by disinhibition of the protein DAAM1 (Dvl associated activator of morphogenesis 1) in cytoplasm. The complex of small GTPase Rac1 and Rho trigger ROCK (Rho kinase) and JNK. Hence, this leads to rearrangements of the cytoskeleton and transcriptional responses. Later, Vangl, a major member of Wnt/PCP signaling cascade is activated via phosphorylation in a Wnt5a-dependent way. Wnt/Ca2+ signaling started by G-protein stimulated phospholipase C activity resulting to intracellular calcium effluxes and downstream calcium dependent cytoskeletal and transcriptional responses (Zhan et al., 2016). The non-canonical Wnt signaling pathway coordinates cell adhesion, cell polarity and tissue motility.(Vincan and Barker, 2008)

Colorectal cancer stem cells (CSCs) and Wnt signaling

There is growing body of evidence showing the a potential link between Wnt pathway and modulation of CSCs (Fodde and Brabletz, 2007). An active Wnt signaling cascade is related with CSC phenotype. Epithelial cellular adhesion molecule (EpCAM), aldehyde dehydrogenase 1-A1 (ALDH1A1), CD24, CD26 (dipeptidyl peptidase IV: DPPIV), CD44, CD133, CD166, Lgr5, and Musashi-1 have been identified as colorectal CSC markers (Lin et al., 2011; Todaro et al., 2010). In particular, Lgr5/orphan G-protein-coupled receptor and CD44 are the well-known TCF/ LEF target genes(Yamamoto et al., 2003). Moreover due to the up regulation of ATP-binding cassette (ABC) family transporters involved in drug-efflux pumps, CSCs often show resistance to conventional

chemotherapeutic regimens. Therefore, it is believed that CSCs are associated with increased risk for cancer relapse post-chemotherapy. ABCB1 is one of the TCF/LEF target genes and have been reported to be involved with intestinal tumorigenesis(Yamamoto et al., 2003).

β -catenin as a therapeutic target for colon cancer

Several agents have been developed for targeting this pathway. It has been shown that retinoic acid could inhibit Wnt signaling by direct interaction with β -catenin/competition for TCF binding (Xiao et al., 2003). Also, the active form of vitamin D (1,25(OH)₂D₃), encourages the β -catenin binding to the vitamin D receptor, consequently decreasing the amount of β -catenin. In addition, 1,25(OH)₂D₃ modulate E-cadherin, which conserves β -catenin in the membrane, preventing its nucleus translocation (Pálmer et al., 2001). The polyphenols curcumin, quercetin, resveratrol and the green tea polyphenol epigallocathechin-3-gallate have been reported to suppress Wnt inhibiting activity (Sarkar et al., 2010). differentiation-inducing factors (DIF) have a phenol unit thus can inhibit β -catenin via activating GSK3-b (Takahashi-Yanaga et al., 2003).

Phytoestrogen and Genistein have suggested to inactivate Wnt signaling by over expression of Ecadherin and GSK3-b. Genistein, a soy-derived isoflavone, plays a role in reversing resistance to platinum and fluoropyrimidines compounds (Su and Simmen, 2009). This agent has been tested in a phase I-II clinical trial in combination with bevacizumab /FOLFOX for metastatic cancer patients. Another natural compounds may have Wnt inhibitor features include 3,3-diindolylmethane ,magnolol, indole-3-carbinol, and lycopene (Sarkar et al., 2010).

Synthetic compounds: NSAIDs

In colon cancer, sulindac and celecoxib have reported to decrease adenomas in FAP patients and celecoxib is FDA approved COX-2 selective inhibitor agent for the treatment of FAP. Celecoxib inhibit Wnt/β-catenin signaling via inducing the TCFs degradation, independent of COX-2(Sebio et al., 2014). However, the emergence of several COX-2 inhibitors, such as Celecoxib, Valdecoxib and

Rofecoxib (Vioxx) has provided the potential in combinational COX-2 inhibitor chemotherapy (Arun and Goss, 2004; Natale, 2003; Sweeney, 2003). Lew and colleagues evaluated the effects of Rofecoxib (Vioxx) and a liposome-mediated APC gene therapy in intestinal neoplasia (Lew et al., 2002). After Vioxx and APC treatment, 70% and 54% reduction were reported in total number of intestinal polyps, respectively, while the combination therapy reduced polyp formation up to 87% (Lew et al., 2002).

Targeted therapies

Several studies have performed for targeting the Wnt cascade components as reported in Table1.

Small molecules

Porcupine is a membrane bound 0-acyltransferase that mediates the palmitoylation of Wnt ligands. Suppression of Porcupine by its inhibitor leads to block of Wnt/ β -catenin signaling activities (Bartscherer and Boutros, 2008). Inhibitors of Wnt production (IWPs) is a novel type of Porcupine-targeted Wnt antagonists. Initially, Chen *et al* discovered IWPs through screening about 200,000 small molecules in L-Wnt-STF cell line using a cell-based Wnt/ β -catenin signal reporter assay. IWPs competitively bound to Porcupine and inhibited Wnt production, while Porcupine overexpressing restored Wnt signaling in cells treated with IWP. Further biochemical researches confirmed that IWPs bind to Porcupine and prevent Wnt palmitoylation, which is substantial for Porcupine -mediated lipidation essential for Wnt secretion (Chen et al., 2009).

LGK974 is another compound which binds and block the porcupine enzyme. LGK974 impedes the growth of murine tumor xenografts model through mouse mammary tumor virus (MMTV)-originated ectopic Wnt1 expression. A phase I open-label clinical trial of LGK974 is ongoing (Liu et al., 2013).

ICG-001 is as antagonist of the protein –protein interaction of β -catenin with CBP(Emami et al., 2004). Suppression of the β -catenin/CBP interaction eradicated tumor-initiating cells and cell differentiation advancement. ICG-001 decrease the growth of colorectal cancer cells in vitro. Antitumor effects of ICG-001 was found in mouse having mutation in one allele of the APC or nude mouse SW620 of colorectal cancer (Eguchi et al., 2005).

PRI-724, the second-generation of CBP/catenin antagonist, was shown to increase p300/β-catenin binding and stem-cell differentiation (Ma et al., 2005). PRI-724, showed acceptable toxicity profile in a phase I clinical trial (Lenz and Kahn, 2014). A phase II trial of combination chemotherapy consisting of bevacizumab alone and with PRI-724 is now being planned for patients with metastatic colorectal cancer.

CWP232291, is another small molecule that inhibited Wnt-mediated transcriptional activity. Active form of CWP232204 binds to Src-Associated substrate in Mitosis of 68 kDa (Sam68) and promotes apoptosis. This agent is being investigated in a phase I trial for hematological cancers (Sebio et al., 2014).

Several small molecules that antagonized the interaction between TCF/LEF1 and β -catenin, have been recognized such as CGP 049090, PNU74654, 2,4 diamino-quinazoline, or PKF115-584(Sebio et al., 2014). Interestingly, common core chemical structure of these compounds indicated that they may have similar binding structure on β -catenin or Tcf and interrupt the Tcf/ β -catenin complex in a similar way(Lepourcelet et al., 2004).

Pyrvinium is an anti-pinworm drug, that is reported to activate the CK1 α kinase and prevent Wnt signaling (Saraswati et al., 2010; Thorne et al., 2010). Other effective small-molecule compounds FJ9(Fujii et al., 2007), NSC668036(Shan et al., 2005), and 3289 –8625(Grandy et al., 2009) target Dv1 PDZ domain (PSD95/Disclarge/Zonula occludens-1) which is necessary for its frizzled interaction (Wong et al., 2003).

Grandy and colleagues identified 3289–8625 (Grandy et al., 2009). They showed that this agent can interact with PDZ domain of Dvl effectively and thereby suppress Wnt3A-induced signaling. (Metcalfe et al., 2010). Recently, salinomycin, an anti-coccidial drug, is detected to target CSCs via suppression of Wnt signaling by LRP6 degradation(Lu and Li, 2014).

Small-molecule XAV939 is a Wnt inhibitor targeting tankyrases 1 and 2 (TNKS1/2)(Huang et al., 2009). TNKS1/2, poly-ADP ribosylate axins and poly-ADP ribosylated axins, are exposing ubiquitination and degradation. In the other words, inhibition of poly-ADP ribosylation consequently leads to stabilization of axins and antagonist Wnt signaling. XAV939 potentially inhibit the proliferation of APC-depletion colorectal cancer cells. Recently, a selective TNKS inhibitor, NVP- TNKS656 was identified through structure-based optimization of XAV939 (Huang et al., 2009).

BBI608 is a small molecule inhibits signal transducer and activator of transcription-3 (Stat3) and targets CSCs (Ciombor et al., 2015a). BBI608 can also suppress β -catenin signaling. A recent phase III trial of BBI608 against metastatic colon cancer was terminated due to loss of expected efficacy in the short-term analysis. At present, BBI608 is under clinical survey in combination with chemotherapeutic drugs such as cisplatin, gemcitabine, paclitaxel, temozolomide, sorafenib, and pemetrexed(Ciombor et al., 2015b).

Monoclonal antibody against frizzled receptors

Vanituctumab (OMP-18R5) is a humanized monoclonal antibody that interact with frizzled receptors and block canonical Wnt signaling in a patient-derived colorectal cancer mouse xenograft model(Gurney et al., 2012).

Decoy receptor

OMP-54 F28 is a recombinant protein of the frizzled 8 cysteine-rich domain that fused to the Fc part of immunoglobulin. OMP-54 F28 competes with frizzled 8 for ligand binding as a decoy

receptor, and then antagonizes Wnt signaling.OMP-54 F28 suppresses tumor growth and inhibits cancer stem cell function (Le et al., 2015). A phase I study of OMP-54 F28 monotherapy is performed in 25 patients with solid tumors. Dysgeusia, fatigue, muscle spasms, decreased appetite, nausea, and vomiting of grades I/II were reported. But, no adverse event was detected (Masuda et al., 2015).

Conclusions and future perspectives

Wnt signaling pathway is one of the key dysregulated pathways in colorectal cancer, which is associated with increased cell proliferation and resistance of the tumor cells to chemotherapy, suggesting its value as a therapeutic target in treatment of CRC. Several agents have been developed to target this pathway and some them are now in clinical phases. However, several questions are still remained to be elucidated.

In aggregate future challenges should focus on the (1) optimization and evaluation of Wnt/b-catenin inhibitors, (2) selection of patient who might benefit from therapy, (3) detection of prognostic and predictive markers that can be used to monitor treatment response; (4) targeting of other key signaling pathways (e.g., AKT/PI3K, NOTCH, mTOR pathway) (Avan et al., 2017; Maftouh et al., 2014; Pashirzad et al., 2016), in parallel, to overcome cell resistance.

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References

Arun B, Goss P. The role of COX-2 inhibition in breast cancer treatment and prevention; 2004. Elsevier. 22-29.

- Barker N. 2008. The canonical Wnt/β-catenin signalling pathway. Wnt Signaling: Pathway Methods and Mammalian Models:5-15.
- Bartscherer K, Boutros M. 2008. Regulation of Wnt protein secretion and its role in gradient formation. EMBO reports 9(10):977-982.
- Chen B, Dodge ME, Tang W, Lu J, Ma Z, Fan C-W, Wei S, Hao W, Kilgore J, Williams NS. 2009. Small molecule– mediated disruption of Wnt-dependent signaling in tissue regeneration and cancer. Nature chemical biology 5(2):100-107.
- Ciombor KK, Edenfield WJ, Hubbard JM, O'Dwyer PJ, Becerra C, Larson T, Spira AI, Grothey A, Manoharan D, Li W. A phase Ib/II study of cancer stem cell inhibitor BBI608 administered with panitumumab in KRAS wild-type (wt) patients (pts) with metastatic colorectal cancer (mCRC) following progression on anti-EGFR therapy; 2015a. p 3617.
- Ciombor KK, Wu C, Goldberg RM. 2015b. Recent therapeutic advances in the treatment of colorectal cancer. Annual review of medicine 66:83-95.
- Eguchi M, Nguyen C, Lee SC, Kahn M. 2005. ICG-001, A Novel Small Molecule Regulator of TCF/β-Catenin Masakatsu Transcription. Medicinal Chemistry 1(5):467-472.
- Emami KH, Nguyen C, Ma H, Kim DH, Jeong KW, Eguchi M, Moon RT, Teo J-L, Oh SW, Kim HY. 2004. A small molecule inhibitor of β-catenin/cyclic AMP response element-binding protein transcription. Proceedings of the National Academy of Sciences of the United States of America 101(34):12682-12687.
- Fodde R, Brabletz T. 2007. Wnt/β-catenin signaling in cancer stemness and malignant behavior. Current opinion in cell biology 19(2):150-158.
- Fujii N, You L, Xu Z, Uematsu K, Shan J, He B, Mikami I, Edmondson LR, Neale G, Zheng J. 2007. An Antagonist of Dishevelled Protein-Protein Interaction Suppresses β-Catenin–Dependent Tumor Cell Growth. Cancer research 67(2):573-579.
- Grandy D, Shan J, Zhang X, Rao S, Akunuru S, Li H, Zhang Y, Alpatov I, Zhang XA, Lang RA. 2009. Discovery and characterization of a small molecule inhibitor of the PDZ domain of dishevelled. Journal of Biological Chemistry 284(24):16256-16263.
- Gurney A, Axelrod F, Bond CJ, Cain J, Chartier C, Donigan L, Fischer M, Chaudhari A, Ji M, Kapoun AM. 2012. Wnt pathway inhibition via the targeting of Frizzled receptors results in decreased growth and tumorigenicity of human tumors. Proceedings of the National Academy of Sciences 109(29):11717-11722.
- Hosseini M. 2016. Exosome-encapsulated microRNAs as potential circulating biomarkers in colon cancer. Current Pharmaceutical Design 22:1-1.
- Huang S-MA, Mishina YM, Liu S, Cheung A, Stegmeier F, Michaud GA, Charlat O, Wiellette E, Zhang Y, Wiessner S. 2009. Tankyrase inhibition stabilizes axin and antagonizes Wnt signalling. Nature 461(7264):614-620.
- Le PN, McDermott JD, Jimeno A. 2015. Targeting the Wnt pathway in human cancers: Therapeutic targeting with a focus on OMP-54F28. Pharmacology & therapeutics 146:1-11.
- Lenz HJ, Kahn M. 2014. Safely targeting cancer stem cells via selective catenin coactivator antagonism. Cancer science 105(9):1087-1092.
- Lepourcelet M, Chen Y-NP, France DS, Wang H, Crews P, Petersen F, Bruseo C, Wood AW, Shivdasani RA. 2004. Small-molecule antagonists of the oncogenic Tcf/β-catenin protein complex. Cancer cell 5(1):91-102.
- Lew JI, Guo Y, Kim RK, Vargish L, Michelassi F, Arenas RB. 2002. Reduction of intestinal neoplasia with adenomatous polyposis coli gene replacement and COX-2 inhibition is additive. Journal of gastrointestinal surgery 6(4):563-568.
- Lin L, Liu Y, Li H, Li P, Fuchs J, Shibata H, Iwabuchi Y, Lin J. 2011. Targeting colon cancer stem cells using a new curcumin analogue, GO-Y030. British journal of cancer 105(2):212-220.
- Liu J, Pan S, Hsieh MH, Ng N, Sun F, Wang T, Kasibhatla S, Schuller AG, Li AG, Cheng D. 2013. Targeting Wntdriven cancer through the inhibition of Porcupine by LGK974. Proceedings of the National Academy of Sciences 110(50):20224-20229.
- Lu W, Li Y. 2014. Salinomycin Suppresses LRP6 Expression and Inhibits Both Wnt/β-catenin and mTORC1 Signaling in Breast and Prostate Cancer Cells. Journal of cellular biochemistry 115(10):1799-1807.
- Ma H, Nguyen C, Lee K-S, Kahn M. 2005. Differential roles for the coactivators CBP and p300 on TCF/β-cateninmediated survivin gene expression. Oncogene 24(22):3619-3631.
- Maftouh M, Belo AI, Avan A, Funel N, Peters GJ, Giovannetti E, Van Die I. Galectin-4 expression is associated with reduced lymph node metastasis and modulation of Wnt/β-catenin signalling in pancreatic adenocarcinoma. Oncotarget. 2014;5(14):5335-49
- Metcalfe C, Ibrahim AE, Graeb M, de la Roche M, Schwarz-Romond T, Fiedler M, Winton DJ, Corfield A, Bienz M. 2010. Dvl2 promotes intestinal length and neoplasia in the ApcMin mouse model for colorectal cancer. Cancer research 70(16):6629-6638.
- Moradi A, Ghasemi F, Anvari K, Hassanian SM, Simab SA, Ebrahimi S, Hesari A, Forghanifard MM, Boroushaki MT, ShahidSales S, Avan A. The Cross-Regulation Between SOX15 and Wnt Signaling Pathway. J Cell Physiol.

2017. doi: 10.1002/jcp.25802.Masuda M, Sawa M, Yamada T. 2015. Therapeutic targets in the Wnt signaling pathway: Feasibility of targeting TNIK in colorectal cancer. Pharmacology & therapeutics 156:1-9.

- Morin PJ, Sparks AB, Korinek V, Barker N, Clevers H, Vogelstein B, Kinzler KW. 1997. Activation of β-catenin-Tcf signaling in colon cancer by mutations in β-catenin or APC. Science 275(5307):1787-1790.
- Natale RB. 2003. Irinotecan, cisplatin/carboplatin, and COX-2 inhibition in small-cell lung cancer. Oncology (Williston Park, NY) 17(7 Suppl 7):22-26.
- Pálmer HG, González-Sancho JM, Espada J, Berciano MT, Puig I, Baulida J, Quintanilla M, Cano A, de Herreros AG, Lafarga M. 2001. Vitamin D3 promotes the differentiation of colon carcinoma cells by the induction of Ecadherin and the inhibition of β-catenin signaling. The Journal of cell biology 154(2):369-388.
- Park S-J, Ahmad F, Philp A, Baar K, Williams T, Luo H, Ke H, Rehmann H, Taussig R, Brown AL. 2012. Resveratrol ameliorates aging-related metabolic phenotypes by inhibiting cAMP phosphodiesterases. Cell 148(3):421-433.
- Pashirzad M, Shafiee M, Rahmani F, Behnam-Rassouli R, Hoseinkhani F, Ryzhikov M, Moradi Binabaj M, Parizadeh MR, Avan A, Hassanian SM. Role of Wnt5a in the Pathogenesis of Inflammatory Diseases. J Cell Physiol. 2016. doi: 10.1002/jcp.25687.
- Preisinger AC, Hedge P, McKechnie D, Finniear R, Markham A, Groffen J, Boguski M, Altschui S, Horii A, Ando H. 1991. Identification of FAP locus genes from chromosome 5q21. Science 253:661-665.
- Saraswati S, Alfaro MP, Thorne CA, Atkinson J, Lee E, Young PP. 2010. Pyrvinium, a potent small molecule Wnt inhibitor, promotes wound repair and post-MI cardiac remodeling. PLoS One 5(11):e15521.
- Sarkar FH, Li Y, Wang Z, Kong D. 2010. The role of nutraceuticals in the regulation of Wnt and Hedgehog signaling in cancer. Cancer and Metastasis Reviews 29(3):383-394.
- Sebio A, Kahn M, Lenz H-J. 2014. The potential of targeting Wnt/β-catenin in colon cancer. Expert opinion on therapeutic targets 18(6):611-615.
- Shan J, Shi D-L, Wang J, Zheng J. 2005. Identification of a specific inhibitor of the dishevelled PDZ domain. Biochemistry 44(47):15495-15503.
- Su Y, Simmen RC. 2009. Soy isoflavone genistein upregulates epithelial adhesion molecule E-cadherin expression and attenuates β-catenin signaling in mammary epithelial cells. Carcinogenesis 30(2):331-339.
- Sweeney CJ. 2003. Why cyclooxygenase-2 inhibition plus chemotherapy? American journal of clinical oncology 26(4):S122-S125.
- Takahashi-Yanaga F, Taba Y, Miwa Y, Kubohara Y, Watanabe Y, Hirata M, Morimoto S, Sasaguri T. 2003. Dictyostelium differentiation-inducing factor-3 activates glycogen synthase kinase-3β and degrades cyclin D1 in mammalian cells. Journal of Biological Chemistry 278(11):9663-9670.
- Thorne CA, Hanson AJ, Schneider J, Tahinci E, Orton D, Cselenyi CS, Jernigan KK, Meyers KC, Hang BI, Waterson AG. 2010. Small-molecule inhibition of Wnt signaling through activation of casein kinase 1α. Nature chemical biology 6(11):829-836.
- Todaro M, Francipane MG, Medema JP, Stassi G. 2010. Colon cancer stem cells: promise of targeted therapy. Gastroenterology 138(6):2151-2162.
- Vincan E, Barker N. 2008. The upstream components of the Wnt signalling pathway in the dynamic EMT and MET associated with colorectal cancer progression. Clinical & experimental metastasis 25(6):657-663.
- Watanabe T, Itabashi M, Shimada Y, Tanaka S, Ito Y, Ajioka Y, Hamaguchi T, Hyodo I, Igarashi M, Ishida H. 2015. Japanese Society for Cancer of the Colon and Rectum (JSCCR) Guidelines 2014 for treatment of colorectal cancer. International journal of clinical oncology 20(2):207-239.
- Wong H-C, Bourdelas A, Krauss A, Lee H-J, Shao Y, Wu D, Mlodzik M, Shi D-L, Zheng J. 2003. Direct binding of the PDZ domain of Dishevelled to a conserved internal sequence in the C-terminal region of Frizzled. Molecular cell 12(5):1251-1260.
- Xiao J-H, Ghosn C, Hinchman C, Forbes C, Wang J, Snider N, Cordrey A, Zhao Y, Chandraratna RA. 2003. Adenomatous polyposis coli (APC)-independent regulation of β-catenin degradation via a retinoid X receptormediated pathway. Journal of Biological Chemistry 278(32):29954-29962.
- Xiao Y, Yang X, Miao Y, He X, Wang M, Sha W. 2016. Inhibition of cell proliferation and tumor growth of colorectal cancer by inhibitors of Wnt and Notch signaling pathways. Oncology Letters 12(5):3695-3700.
- Yamamoto H, Ihara M, Matsuura Y, Kikuchi A. 2003. Sumoylation is involved in β-catenin-dependent activation of Tcf-4. The EMBO journal 22(9):2047-2059.
- Zhan T, Rindtorff N, Boutros M. 2016. Wnt signaling in cancer. Oncogene. 2016. doi: 10.1038/onc.2016.304

Table 1. Summary of	of Wnt-β-catenin pathwa	y inhibitors in clinical	development			
Compound	Molecular target	Tumor	Trial	Arms	Status	Trial identifier
name			Phase			
LGK974	Inhibitor of	BRAF mutant	I/II	WNT974+	not yet	NCT02278133
(WNT974)	Porcupine	colorectal cancer		LGX818+Cetuximab	recruiting	
LGK974	Inhibitor of	BRAF Mutant	Ι	LGK974	suspended	NCT01351103
(WNT974)	Porcupine	Colorectal Cancer			participant	
					recruitment	
Foxy-5	Wnt5a mimetic	colorectal cancer	Ι	Foxy-5	recruiting	NCT02655952
					participants	
Foxy-5	Wnt5a mimetic	Colorectal Cancer	Ι	Foxy-5	terminated	NCT02020291
Vanituctumab	Frizzled $-1, -2, -5,$	Solid tumors	Ι	OMP-18R5	completed	NCT01345201
(OMP-18R5)	-7, -8 receptors					
OMP-54 F28	Frizzled-8 receptor	Solid tumors	Ι	OMP-54 F28	completed	NCT01608867
(Decoy receptor)	-				-	
Niclosamide	Frizzled receptor	Colon Cancer	Ι	Niclosamide	not yet	NCT02687009
					recruitment	
OMP131R10	Anti-R-spondin3	RSPO3 biomarker-	Ι	OMP-131R10	recruiting	NCT02482441
	antibody	positive metastatic		FOLFIRI	participants	
	-	colorectal				
		cancer				
PRI-724	Interaction of	Advanced solid	Ι	PRI-724	terminated	NCT01302405
	β-catenin and CBP	tumors				
PRI-724	Interaction of	Metastatic	Π	PRI-724	not yet	
	β-catenin and CBP	Colorectal Cancer		+mFOLFOX6+bevacizumab	recruitment	NCT02413853
Genistein	GSK3- b	Metastatic	I/II	Genistein	recruiting	NCT01985763
-		Colorectal Cancer			participants	
Aspirin	COX-2	Colorectal	Π	Aspirin	not yet	NCT02965703
-		Adenoma		Placebo	recruitment	
BBI608	Unknown	Metastatic colon	Ι	BBI608	recruiting	NCT02641873
		cancer		+FOLFIRI+Bevacizumab	participants	
BBI608	Unknown	Metastatic colon	Ib/II	BBI608 + Pembrolizumab	recruiting	NCT02851004
		cancer			participants	
BBI608	Unknown	Metastatic colon	III	BBI-608 + Leucovorin+	recruiting	NCT02753127
		cancer		Irinotecan+Bevacizumab	participants	
				Fluorouracil +Leucovorin+		
				Irinotecan+Bevacizumab		
Curcumin	Tcf/β-catenin	Colorectal	II	Anthocyanins +Phospholipidic	recruiting	NCT01948661
	complex	Adenoma		Curcumin	participants	
	-			Placebo	_	
Resveratrol	PDE4	Colon cancer	Ι	Resveratrol	completed	NCT00256334